

17th Jan 2018, Mid term review and Research symposium

**Updates:
Research on cassava witches' broom
disease and cassava mosaic disease**

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International Center for Tropical Agriculture
Since 1967 Science to cultivate change

Overview of CMD and CWBD

Cassava witches' broom disease



Cassava mosaic disease



First Report in SEA

1990s in TH

2015 in KH

Occurrence

VN, KH, LA, TH, ID, PH, MM, ...

KH and VN

Yield loss

Approx. 34%

Unknown

Causal pathogen

Bacterium (phytoplasma)

Virus

Diagnostics

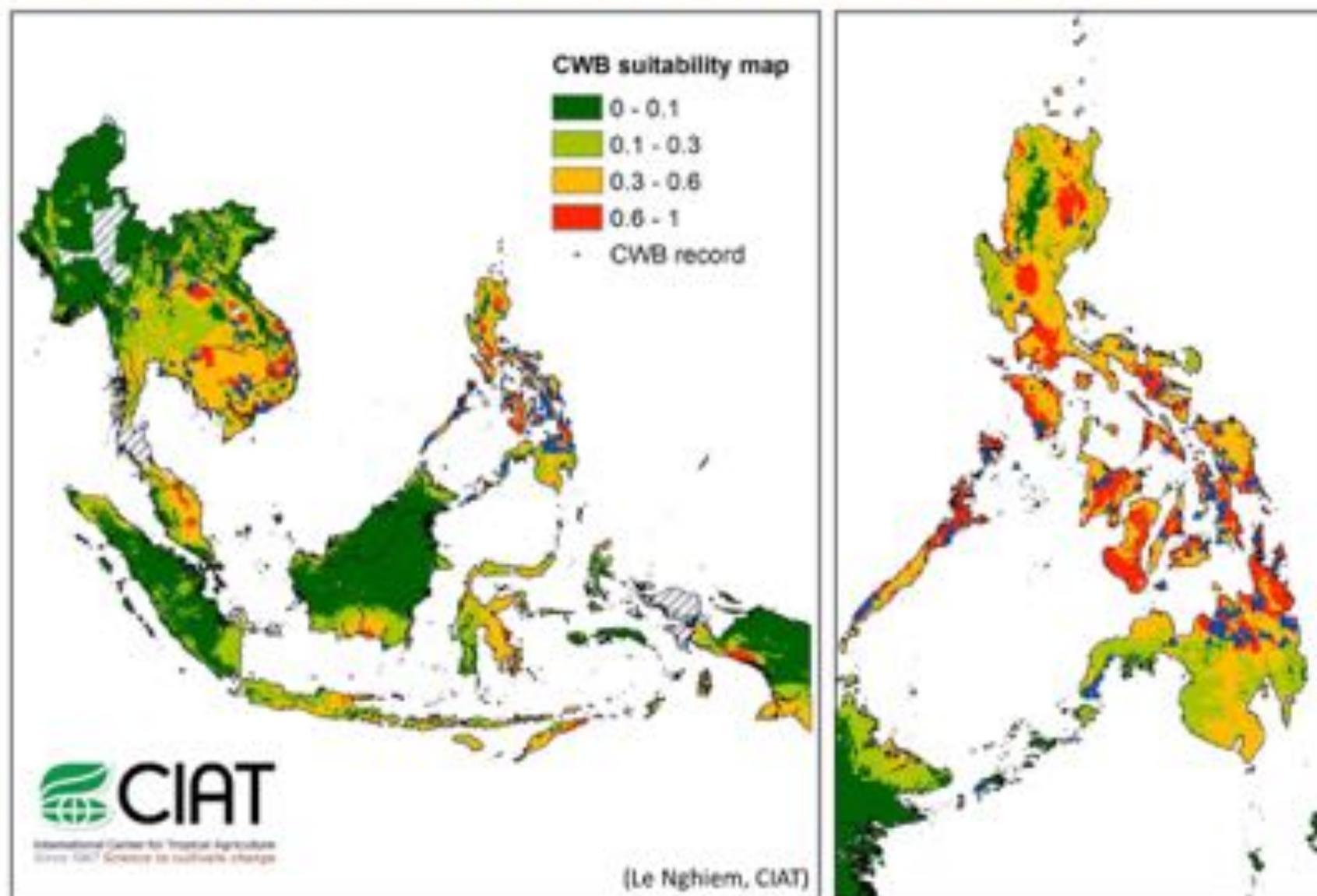
Nested PCR, LAMP, SMART-DART

PCR, ELISA, LAMP



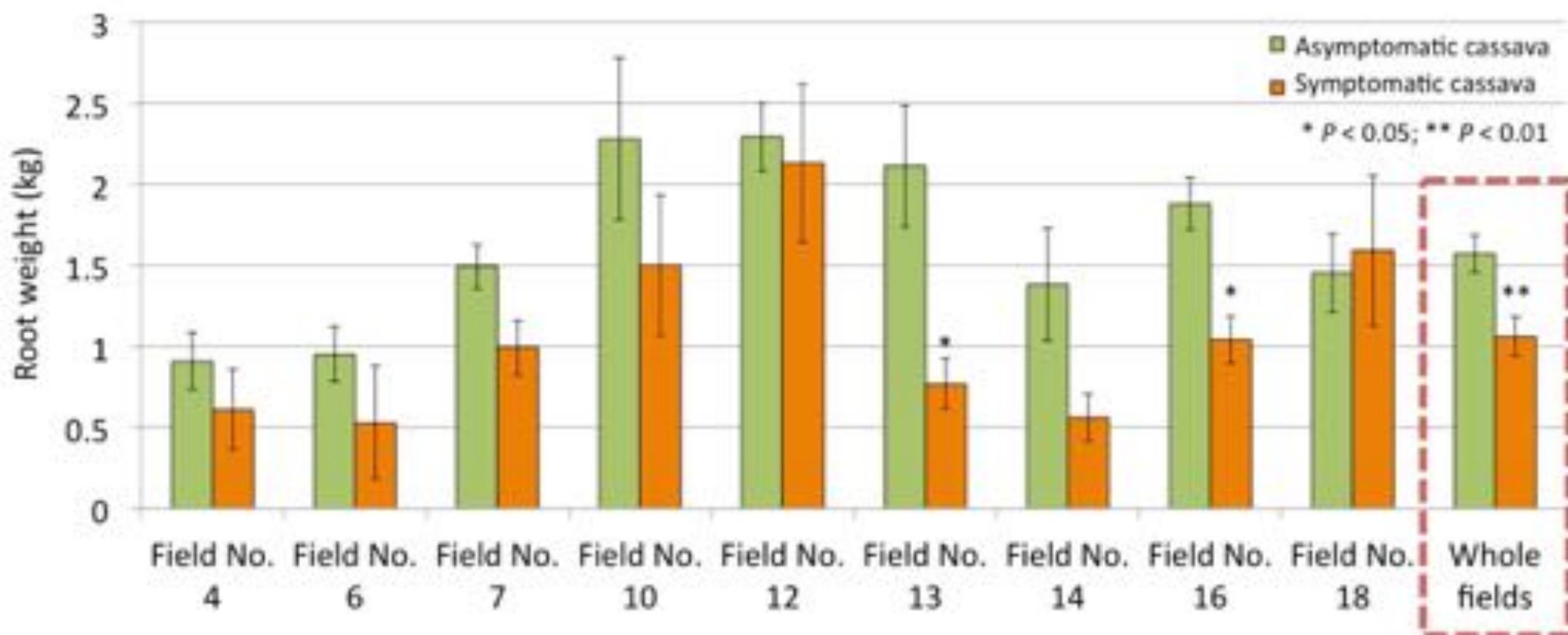
Cassava witches' broom disease

Where CWB occurred, and is exposed to risks



Cassava witches' broom disease

Yield Reduction



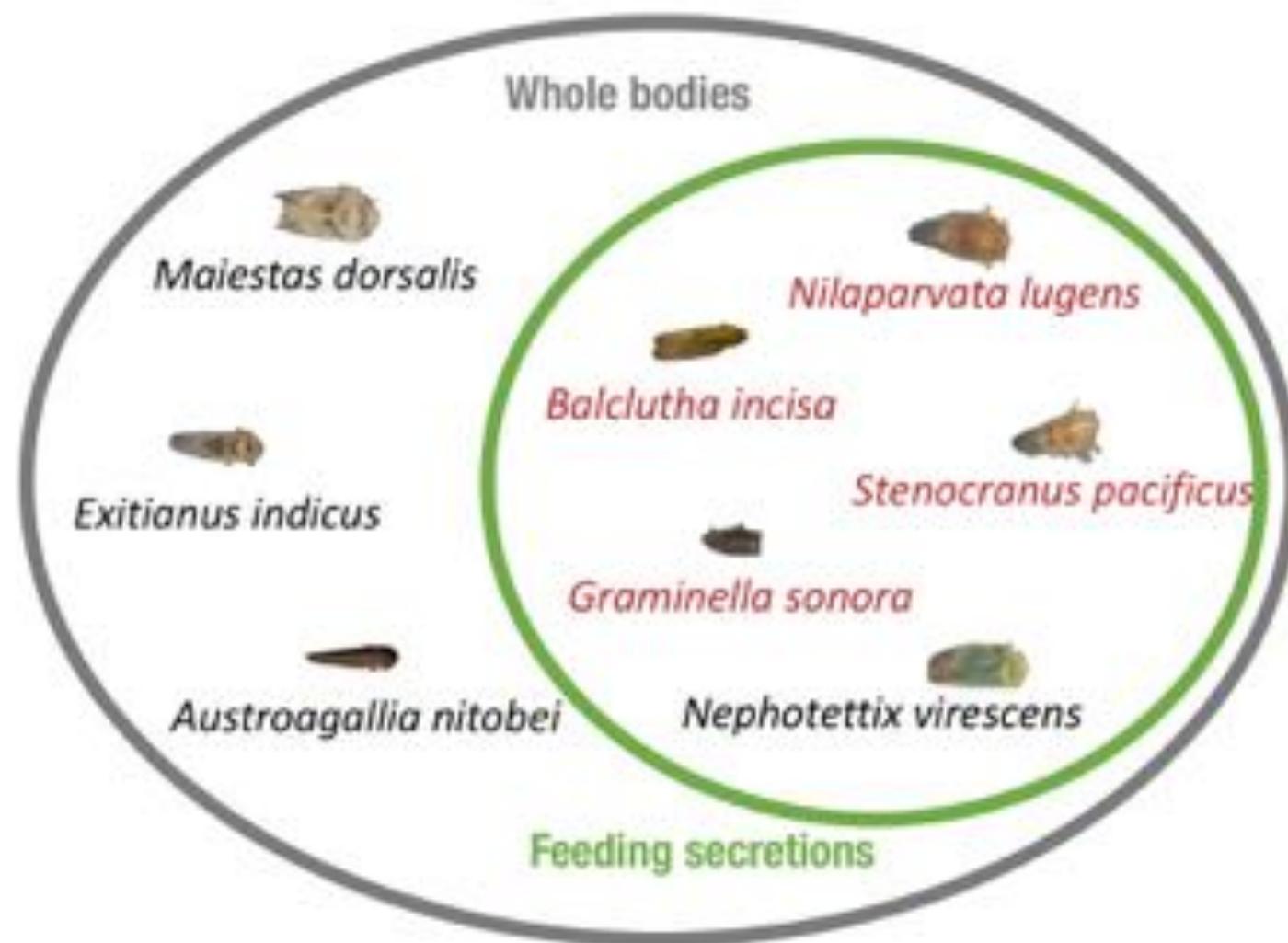
CWB causes yield reduction of approx. 34%



Cassava witches' broom disease

Research on vector insects

Species retaining phytoplasma in cassava fields



Vector candidates

For conclusive evidence, we will conduct *in planta* transmission trials



'Resistance' to phytoplasma?

- Phytoplasma is unculturable
- Resistance to phytoplasma has not commonly been found
- No curative approaches available

High potential for phytoplasma control

- Stable access to clean seeds
- Vector management



Importance and needs of P&D research

Diagnostics

Capacity development of basic diagnostics

Development of diagnostic tools for fields/ local institutions

Monitoring

Systematic sequential monitoring in broader area

Variety evaluation

No resistance (CWB), both CWB and CMD are stake-borne

→ Establishment of seed systems is important

Systematic variety evaluation (eg. yield loss) combined with fingerprinting



Thank you



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Science for a Food Secure Future



Cassava witches' broom

Cassava Witches' Broom (CWB) : An emergent disease in SE Asia

- CWB is thought to be caused by phytoplasma
- Presence in Asia and Central/ Latin Americas
- In Africa: Arocha *et al.*, 2009

Countries	Reported causal agent	References
Vietnam	group 16Srl & II	Alvarez <i>et al.</i> , 2013; Trinh <i>et al.</i> , 2015
Brazil	group 16SrIII	Flores <i>et al.</i> , 2013
Cuba	group 16Srl	Arocha <i>et al.</i> , 2009b
Uganda	group 16SrII	Arocha <i>et al.</i> , 2009a



(Photo by Georgina Smith / CIAT)

Table 1. Main arthropod taxa reported in SE Asian cassava crops, organised by year of first (regional) record (taxa reported within the past 15 years are highlighted in bold)^a

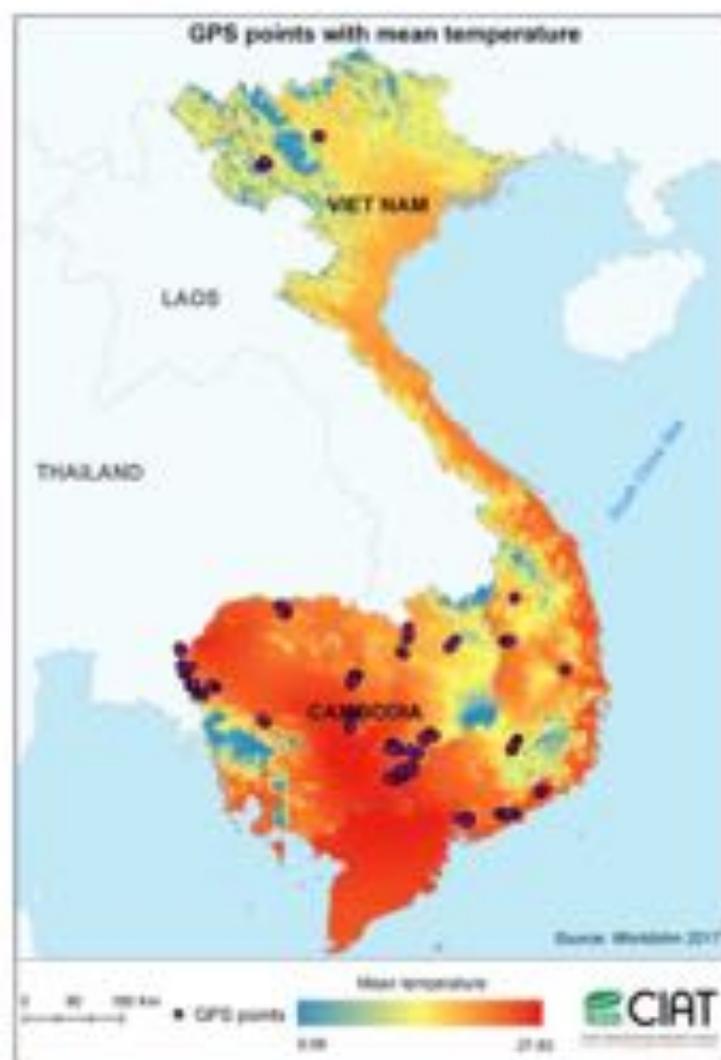
Taxa	Common name	Origin	Record SE Asia	Maximum yield loss(%) ^b	References
A. Short-tailed mealybug					
<i>Phenacoccus manihoti</i>	Cassava mealybug	Neotropical	2008	50,* 84**	19
<i>Paracoccus marginatus</i>	Papaya mealybug	Nearctic	2008	40***	17,203
<i>P. solenopsis</i>	Cotton mealybug	Nearctic	2006		34,42
<i>P. madeirensis</i>	Madeira mealybug	Neotropical	2004	n.a.	16,34
<i>Maconellicoccus hirsutus</i>	Hibiscus mealybug	Neotropical	1942		43,204
B. Long-tailed mealybug					
<i>Pseudococcus jackbeardsleyi</i>	Jack Beardsley mealybug	Neotropical	1996	n.a.	16,34
<i>Femisia virgata</i>	Striped mealybug	Neotropical	1942		43,45
C. Green mite complex					
<i>Mononychellus mcgregori</i>	Green mite	Neotropical	2009	60****	21,61
D. Red mite complex					
<i>Neotetranychus lek</i>	Red mite	n. a.	2013		59
<i>Oligonychus thelytokus</i>	Red mite	Afrotropical	1998		53,205
<i>Tetranychus marianae</i>	Red mite	Nearctic	1975		55,56
<i>T. just</i>	Red mite	Neotropical	1975		49,55
<i>T. urticae</i>	Red mite	Palaearctic	1975		49,55
<i>Eutetranychus africanus</i>	African red mite	Afrotropical	1975		49,56
<i>O. biharensis</i>	Red mite	Oriental	1975		49,56
<i>T. cinnabarinus</i>	Red mite	Palaearctic	1969	53*****	49,54
<i>T. neocaledonicus</i>	Red mite	Australasian	1962		49,206
<i>T. truncatus</i>	Red mite	Palaearctic	1962		49,206
<i>T. karzowii</i>	Red mite	Palaearctic	1962		49,206
<i>E. orientalis</i>	Red mite	Palaearctic	1962		49,206
E. Whiteflies					
<i>Aleurodicus dispersus</i>	Spraying whitefly	Nearctic	1987		21,70
<i>Bemisia tabaci</i>	Silverleaf whitefly	Palaearctic	1933	n.a.	21,75
F. Scale insects					
<i>Aonidomytilus albus</i>	Cassava scale	Nearctic	1935		19,73
<i>Parasaissetia nigra</i>	Black scale	n. a.		n.a.	53
<i>Saissetia miranda</i>	Black scale	Nearctic	n. a.		53
G. Others					
Isoptera: Rhinotermitidae	Termites	Oriental	n. a.	n.a.	19,53
Coleoptera: Scarabaeidae	Whitegrubs	Oriental	n. a.	n.a.	94,87

^a n. a. = information not available.



Supplementary data

Surveyed districts



Location of 419 field sampling sites in Cambodia & Vietnam.

- Covering big production areas
- 15 districts in each country

Country	District
Vietnam	1 Thuan Chau
	2 Van Yen
	3 Chu Prong
	4 Mad Lak
	5 Eaker
	6 Dak Glong
	7 Sa Thay
	8 Krong Pa
	9 Song Hinh
	10 Bac Binh
	11 Ham Tan
	12 Tan Bien
	13 Tan Chau
	14 Ham Thuan Nam
	15 Long Thanh

Country	District
Cambodia	1 Anlong Veaeng
	2 Malai
	3 Sala Krau
	4 Palin
	5 Kamrieng
	6 Phnum Proek
	7 Rattanak mondul
	8 Kravanh
	9 Koun Morn
	10 Steung Treng
	11 Snoul
	12 Dambae
	13 Memot
	14 Romeas Hark
	15 Sandan
	16 Baray



What Phytoplasma is

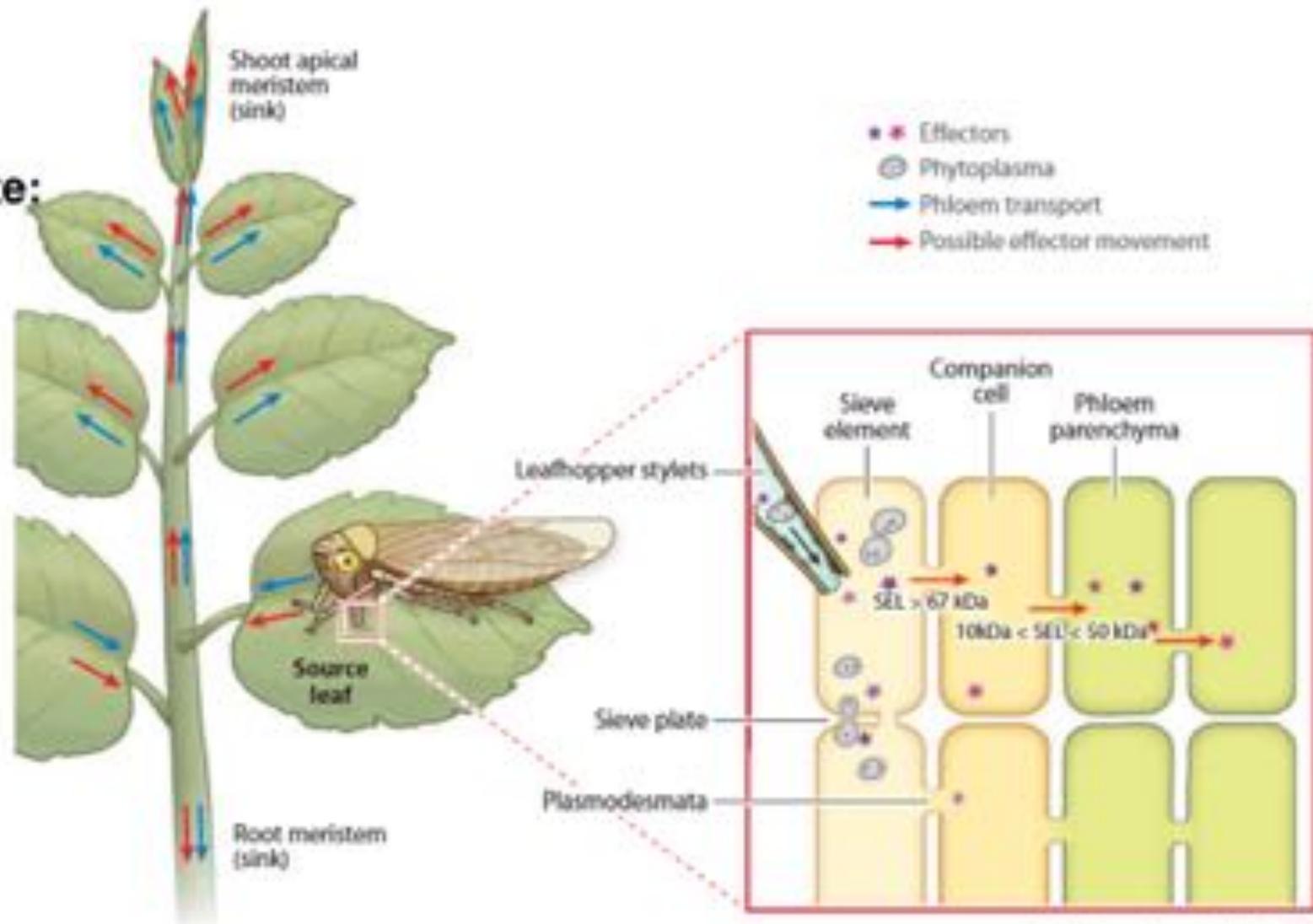
- Plant-pathogenic bacterium





What Phytoplasma is

- Plant-pathogenic bacterium
- Intracellular parasite: phloem-restricted**
- Cell wall-less
- Until now, unable to be cultured



(Sugio *et al.*, 2011)



What Phytoplasma is

- Plant-pathogenic bacterium
- Intracellular parasite: phloem-restricted
- Cell wall-less
- Until now, unable to be cultured
- **Transmitted by insects & parasitic plants**



Insects (Hemiptera)

<http://www.quepasa.com/ky/West-Pennant-hydrich>



Dodder (*Cuscuta* spp.)

<https://islandstrust.wordpress.com/2014/08/img-0165.jpg>





What Phytoplasma is

- Plant-pathogenic bacterium
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Insects (Hemiptera)

<http://www.quepasa.com/ky/West-Pennsylvania>



Dodder (*Cuscuta* spp.)

<https://islandstrust.wordpress.com/2014/08/img-0165.jpg>

- Cicadellidae (Leafhoppers; 叶蝉)
- Delphacidae (Planthoppers; 飛蝨)
- Psyllids



Objectives

Why identify vectors?



<http://www.istock.com/174486888/kyche>

- Phytoplasma is unculturable
- Resistance to phytoplasma has not been found
- No curative approaches available



- **Vector management**
= high potential for phytoplasma control
- Facilitates resistance screening
- Sheds light upon disease epidemiology
- Constitutes basis for vector management



Conclusion

